

Microdialysis - principles of recovery

Recovery - definition of terms

The dialysing properties of the microdialysis probe can be expressed as its recovery for a particular substance.

By comparing the concentration of the substance in the microdialysis probe effluent with the concentration of the medium it is possible to calculate the recovery of the substance.

Relative recovery will approach 100% as the flow rate approches zero, and decrease as the flow rate increases. It is commonly expressed in percent.

Absolute recovery is defined as the mass of a substance recovered during a defined time period. It is zero when the flow rate is zero, and will reach a maximum at higher flow rates, as shown in Fig. 1.

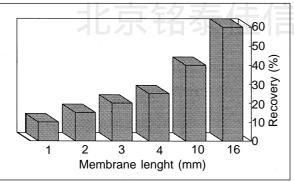


Fig.2. Effect of membrane length on recovery Experimental conditions: CMA/12 Microdialysis Probes Temperature: +20 ° C Flow rate 2 µl/min Test substance: Dopamine

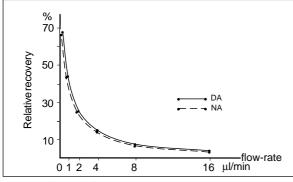


Fig.3. Effect of flow rate on recovery Experimental conditions: CMA/12 Microdialysis **Probe** with 4 mm membrane, Temperature: +20 ° C Test substances: Dopamine, Noradrenaline

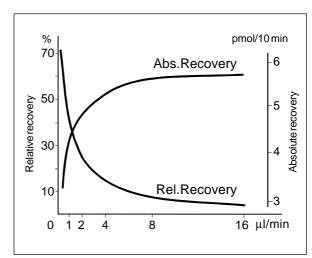


Fig.1. The relative and absolute recoveries for Dopamine as a function of flow rate. CMA/10 Microdialysis Probe, 4 mm membrane.

The recovery of substances in vitro is depending upon:

- length and diameter of the membrane (larger area - higher recovery)
- flow rate
- temperature
- molecular weight of the substance
- molecular shape of the substance
- molecular charge
- binding to the membrane and tubing

Other factors such as pH of the medium and degradation of the substance may also affect the recovery.

Before measuring endogenous substances, it is always advisable to perform an in vitro experiment, to establish the dialysing properties of the microdialysis probe for the particular substance(s) of interest.

It is also possible to study the condition of the probe from day to day by in vitro test.

Recovery experiment

Materials:

CMA/10 Microdalysis Probe 4 mm membrane CMA/100 Microinjection Pump CMA/130 In Vitro Stand CMA/140 Microfraction Collector Microsyringe 1 ml

Perfusion liquid: Ringer´s solution 147 mM Na $^+$, 2.4 mM Ca $^{2+}$, 4 mM K $^+$, 155.6 mM Cl-, pH 6.0

Medium: Catecholamines, 10-6M

Three samples were collected from the microdialysis probe effluent, in $300\,\mu l$ polyethylene vials in 10 min fractions, and compared with samples taken from the medium.

The samples were analyzed by liquid chromatography with electrochemical detection. See Fig. 4.

The medium consisted of catecholamines dissolved in Ringer, at a concentration of $10^{-6} \rm M$ in an Eppendorf vial in the CMA/130 In Vitro Stand.

The CMA/10 Microdialysis Probe was prepared according to the instruction in the package. The probe was perfused with 2 μ l/min. for 30 min. to equlibrate the system before starting the collection of samples.

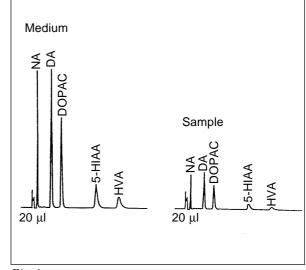


Fig.4.

In Vitro recovery +20 °C

| Substance 2mm 3mm 4mm Dopamine DOPAC 14% 19 24 5-HT 13 17 24 5-HIAA 14 17 24 HVA 13 19 24 Noradrenaline 14 18 23 Na* 30 39 49 K* 48 61 71 Pyruvate 26 38 45 Lactate 23 34 39 Hypoxanthine 17 23 31 |
|---|
| DOPAC 15 19 24 5-HT 13 17 24 5-HIAA 14 17 24 HVA 13 19 24 Noradrenaline 14 18 23 Na+ 30 39 49 K+ 48 61 71 Pyruvate 26 38 45 Lactate 23 34 39 Hypoxanthine 17 23 31 |
| 5-HT 13 17 24 5-HIAA 14 17 24 HVA 13 19 24 Noradrenaline 14 18 23 Na+ 30 39 49 K+ 48 61 71 Pyruvate 26 38 45 Lactate 23 34 39 Hypoxanthine 17 23 31 |
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| Na* 30 39 49 K* 48 61 71 Pyruvate 26 38 45 Lactate 23 34 39 Hypoxanthine 17 23 31 |
| K+ 48 61 71 Pyruvate 26 38 45 Lactate 23 34 39 Hypoxanthine 17 23 31 |
| Pyruvate 26 38 45 Lactate 23 34 39 Hypoxanthine 17 23 31 |
| Lactate 23 34 39 Hypoxanthine 17 23 31 |
| Hypoxanthine 17 23 31 |
| 1 = |
| |
| Inosine 11 17 21 |
| Guanosine 12 16 21 |
| Adenine 15 21 29 |
| Adenosine 10 14 19 |
| Glucose 13 18 22 |
| Acetylcholine 17 24 27 |
| Choline 20 26 31 |
| Aspartate 13 16 24 |
| Asparagine 14 17 25 |
| Glutamate 13 16 24 |
| Serine 12 14 22 |
| Glutamine 14 16 24 |
| Taurine 18 22 27 |
| Tyrosine 14 16 20 |
| GABA 17 21 25 |
| α-aminobutyric acid 16 20 30 |
| Tryptophan 14 19 27 |
| Methionine 14 18 27 |
| Valine 14 18 26 |
| Phenylalanine 14 18 26 |
| Isoleucine 13 18 26 |
| Leucine 14 19 26 |
| Insulin 4 |
| Glucagon 1 |
| Somatostatin 6 |
| ۷۱۲ |

| | Membrane length | |
|------------------|-----------------|------|
| Substance | 2mm | 5mm |
| Angiotensin | 9.4% | 19 |
| AVP | 9.1 | 18.3 |
| β-Endorphin | 1.4 | 3 |
| Bombesin | 8.1 | 16.6 |
| CCK-8 | 6.2 | 12.7 |
| CRF | 1.6 | |
| Dynorphin 1-17 | 3.3 | 6.5 |
| LHRH | 8 | 15.6 |
| [Leu]enkephalin | 10.5 | 20.9 |
| [Met]enkephalin | 13 | 24.8 |
| Neurotensin | 6.3 | 12 |
| NPY | 0.7 | 1.5 |
| Oxytocin | 8.6 | 16.4 |
| Substance K | 9.1 | 18 |
| Substance P | 7.5 | 15.5 |
| TRH | 11.8 | 19.4 |
| | | |

Tab.1. In vitro recovery. Flow rate 2 μl/min. Temperature: +20 ° C

Molecular weight and recovery

Linear relationship between the \log % recovery (in vitro) shown by a CMA/10 Microdialysis Probe and the molecular weight of the substance sampled indicates an exponential relationship between these two factors.

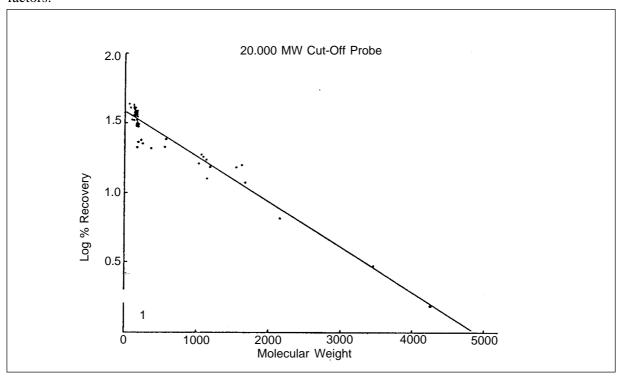


Fig. 5. Recoveries of 42 different substances are plotted. Recovery is minimal at approximately 5000 MW, even though the membrane's nominal cut-off is 20 000. CMA/10 Microdialysis Probe, 5 mm membrane. Flow rate 2 μ l/min.

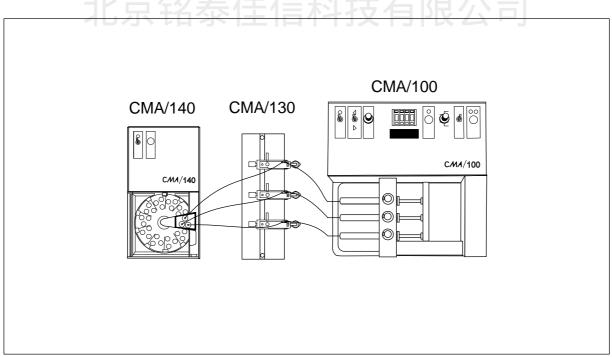


Fig. 6. A typical setup for an in vitro recovery experiment.

(Data on peptides in Tab. 1. and Fig. 5. kindly provided by Dr. K. Kendrick, A.F.R.C. Institute on Animal Physiology and Genetics Research, Babraham, Cambridge U.K.)

| If you require further details on Microdia tion or bibliography, please do not hesitat | lysis procedures, HPLC analysis, instrumentae to contact: |
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